

Volume 11 Number 34, 10 September, 2017 ISSN 1996-0875



ABOUT JMPR

The Journal of Medicinal Plant Research is published weekly (one volume per year) by Academic Journals.

The Journal of Medicinal Plants Research (JMPR) is an open access journal that provides rapid publication (weekly) of articles in all areas of Medicinal Plants research, Ethnopharmacology, Fitoterapia, Phytomedicine etc. The Journal welcomes the submission of manuscripts that meet the general criteria of significance and scientific excellence. Papers will be published shortly after acceptance. All articles published in JMPR are peer reviewed. Electronic submission of manuscripts is strongly encouraged, provided that the text, tables, and figures are included in a single Microsoft Word file (preferably in Arial font).

Contact Us

Editorial Office: jmpr@academicjournals.org

Help Desk: helpdesk@academicjournals.org

Website: http://www.academicjournals.org/journal/JMPR

Submit manuscript online http://ms.academicjournals.me/

Editors

Prof. Akah Peter Achunike

Editor-in-chief
Department of Pharmacology & Toxicology
University of Nigeria, Nsukka
Nigeria

Associate Editors

Dr. Ugur Cakilcioglu

Elazig Directorate of National Education Turkey.

Dr. Jianxin Chen

Information Center,
Beijing University of Chinese Medicine,
Beijing, China
100029,
China.

Dr. Hassan Sher

Department of Botany and Microbiology, College of Science, King Saud University, Riyadh Kingdom of Saudi Arabia.

Dr. Jin Tao

Professor and Dong-Wu Scholar,
Department of Neurobiology,
Medical College of Soochow University,
199 Ren-Ai Road, Dushu Lake Campus,
Suzhou Industrial Park,
Suzhou 215123,
P.R.China.

Dr. Pongsak Rattanachaikunsopon

Department of Biological Science, Faculty of Science, Ubon Ratchathani University, Ubon Ratchathani 34190, Thailand.

Prof. Parveen Bansal

Department of Biochemistry
Postgraduate Institute of Medical Education and
Research
Chandigarh
India.

Dr. Ravichandran Veerasamy

AlMST University
Faculty of Pharmacy, AlMST University, Semeling - 08100,
Kedah, Malaysia.

Dr. Sayeed Ahmad

Herbal Medicine Laboratory, Department of Pharmacognosy and Phytochemistry, Faculty of Pharmacy, Jamia Hamdard (Hamdard University), Hamdard Nagar, New Delhi, 110062, India.

Dr. Cheng Tan

Department of Dermatology, first Affiliated Hospital of Nanjing Univeristy of Traditional Chinese Medicine. 155 Hanzhong Road, Nanjing, Jiangsu Province, China. 210029

Dr. Naseem Ahmad

Young Scientist (DST, FAST TRACK Scheme)
Plant Biotechnology Laboratory
Department of Botany
Aligarh Muslim University
Aligarh- 202 002,(UP)
India.

Dr. Isiaka A. Ogunwande

Dept. Of Chemistry, Lagos State University, Ojo, Lagos, Nigeria.

Editorial Board

Prof Hatil Hashim EL-Kamali

Omdurman Islamic University, Botany Department, Sudan.

Prof. Dr. Muradiye Nacak

Department of Pharmacology, Faculty of Medicine, Gaziantep University, Turkey.

Dr. Sadiq Azam

Department of Biotechnology, Abdul Wali Khan University Mardan, Pakistan.

Kongyun Wu

Department of Biology and Environment Engineering, Guiyang College, China.

Prof Swati Sen Mandi

Division of plant Biology, Bose Institute India.

Dr. Ujjwal Kumar De

Indian Vetreinary Research Institute, Izatnagar, Bareilly, UP-243122 Veterinary Medicine, India.

Dr. Arash Kheradmand

Lorestan University, Iran.

Prof Dr Cemşit Karakurt

Pediatrics and Pediatric Cardiology Inonu University Faculty of Medicine, Turkey.

Samuel Adelani Babarinde

Department of Crop and Environmental Protection, Ladoke Akintola University of Technology, Ogbomoso Nigeria.

Dr.Wafaa Ibrahim Rasheed

Professor of Medical Biochemistry National Research Center Cairo Egypt.

Journal of Medicinal Plants Research

Table of Contents: Volume 11 Number 34 10 September, 2017

ARTICLE

Preliminary phytochemical analysis and the effect of *Agave sisalana* on body weight and defensive behaviours in ovariectomized rats

Telma Gonçalves Carneiro Spera de Andrade, Yara Emilia Arlindo da Silva, Diego Gazoni Espinoza, Leonardo Mateus de Lima, Alessandra Karla de Oliveira Cezar, Isabella Gonçalves Carneiro Spera de Andrade, Pedro de Oliva Neto, Julia Lainer Palacios and Lucinéia dos Santos 538

academicJournals

Vol. 11(34), pp. 538-548, 10 September, 2017

DOI: 10.5897/JMPR2017.6382 Article Number: 0E0D5A066037

ISSN 1996-0875 Copyright © 2017 Author(s) retain the copyright of this article http://www.academicjournals.org/JMPR **Journal of Medicinal Plants Research**

Full Length Research Paper

Preliminary phytochemical analysis and the effect of Agave sisalana on body weight and defensive behaviours in ovariectomized rats

Telma Gonçalves Carneiro Spera de Andrade^{1*}, Yara Emilia Arlindo da Silva¹, Diego Gazoni Espinoza¹, Leonardo Mateus de Lima¹, Alessandra Karla de Oliveira Cezar¹, Isabella Gonçalves Carneiro Spera de Andrade¹, Pedro de Oliva Neto², Julia Lainer Palacios³ and Lucinéia dos Santos³

¹Laboratory of Physiology, Department of Biological Sciences, School of Sciences, São Paulo State University (Unesp), Assis, SP, Brazil.

²Laboratory of Industrial Biotechnology, Department of Biotechnology, School of Sciences, São Paulo State University (Unesp), Assis, SP, Brazil.

³Laboratory of Pharmaceutical Technology in Phytotherapy, Department of Biotechnology, School of Sciences, São Paulo State University (Unesp), Assis, SP, Brazil.

Received 31 March, 2017; Accepted 21 August, 2017

Agave sisalana is a plant belonging to the Agavaceae family. Sisal juice is constituted of steroidic saponins which are precursor molecules of many pharmacologically active steroids. These precursor molecules can act in brain structures that are related to the modulation of emotional disorders. They can act on energy metabolism or directly on the absorptive process of fats. The objective of this research was to identify the biocompounds present in A. sisalana juice, and to evaluate the effect of different extracts on the expression of defensive behaviours of rats in elevated plus maze and open field tests, and body weight, in a condition that corresponds to an induced menopause. Wistar rats were subjected to bilateral ovariectomy or to a sham surgical procedure under anaesthesia. Following surgery, they were treated gavage with sisal juice - concentrated crude juice (CCJ): 500 or 1000 mg/kg; dried extract (DE): 50, 100 or 200 mg/kg; dry mucilage extract (DME): 25, 50 or 100 mg/kg; and intermediate product (IP): 25, 50 or 100 mg/kg; or distilled water. The results showed the presence of coumarins, flavonoids, condensed tannins, free anthraquinones, alkaloids and saponins in the sisal juice. Both CCJ and DE (100 mg/kg) caused weight loss without alteration of defensive behaviours related to the manifestation of anxiety. As saponins were identified in DE in significant amounts, the observed effects were attributed to this component. Such findings point to A. sisalana as a plant that could potentially be used to treat weight gain during menopause.

Key words: Agave sisalana, steroidic saponins, body weight, anxiety, plus-maze, females, ovariectomy.

INTRODUCTION

Agave sisalana is a plant belonging to the family Agavaceae, and it is popularly known in Brazil as sisal (Martin et al., 2009). The fibre is the main product of A.

sisalana culture; it is used in the automotive industry and in the manufacture of ropes, twines, sea cables, carpets, bags, brooms, upholstery and crafts (Silva and Beltrão,

1999; Martin et al., 2003). The fibre-loosening process generates about 95% waste, which corresponds to the watery part of the plant and the bagasse (Pizarro et al., 1999). When not disposed of, the fibre is used as animal feed, biofertilisers and in the production of drugs, particularly hormones (Martin et al., 2009; Ribeiro et al., 2013).

In an ethnopharmacological approach, the juice of *A. sisalana* (that is, the liquid residue of the mucilage) is applied topically to treat skin diseases (El-Hilaly et al., 2003). It has also been administered orally for the treatment of indigestion, bloating, jaundice, constipation and diarrhoea (Bown, 1995); as an analgesic (Duke and Ayensu, 1985); and as a uterine stimulant (Sharaf and Zahran, 1967).

Several organic compounds have been isolated that make up sisal juice, such as oxalic acid, cortisone and saponins (Azevedo et al., 2003). The structure of saponins consists of a glycidic portion linked to an aglycone portion called a sapogenin; in a steroidal skeleton structure it is called steroidal sapogenin (Hostettmann and Marston, 1995; Simons et al., 2006). Sapogenins that are found in A. sisalana are tigogenin, diosgenin and hecogenin (Pizarro et al., 1999). The steroidal saponins are of considerable economic importance as precursors to many pharmacologically active steroids (Oashi, 1999). The use of different saponins, for example, Panax ginseng (Attele et al., 2002), Panax japonicas (Yun, 2010) and Platycodi radix (Han et al., 2000; Zhao et al., 2005) has been validated in different models to prevent or decrease obesity. Studies report that saponins might be able to interfere with the metabolism of cholesterol and fat absorption by inhibiting pancreatic lipase activity (Dickel et al., 2007). In addition, it has been reported that treatment with saponins that were isolated from the plant Panax guinguefolium caused an anxiolytic effect in the elevated plus maze (EPM), among other tests (Wei et al., 2007). These findings indicate a possible relationship between the presence of saponins that were identified in A. sisalana and effects such as weight loss and reduced anxiety.

Periods during which sex steroids in women are found in low concentrations in the serum have been found to be correlated with signs of anxiety, depression and other physiological changes. During the climacteric, the transition phase from the reproductive to non-transition phase from the reproductive to non-reproductive phases of life, which is part of the ageing process in women, often results in hot flashes or hot flushes (Sclowitz et al., 2005), high levels of cholesterol and an increase in body weight (Toth et al., 2000). The accumulation of weight during menopause, especially in the abdominal region, is related to an increase in cardiovascular diseases,

diabetes mellitus type 2, cancer and knee osteoarthritis (Milewicz and Jedrzejuk, 2006; Popov et al., 2007). Metabolic and hormonal changes could be responsiblefor considerably damage in the psychosocial aspects, leading to a reduction in quality of life (Silva et al., 2003).

Considering that the hormone replacement therapy (HRT) have controversies, and their use is contraindicated in patients at risk for some types of cancer and other pathologies (Lima and Baracat, 1995; Bonduki et al., 2006), several studies have been conducted to analyse the therapeutic potential of plants that have phytoestrogens, in the treatment of symptoms of menopause (Rachev et al., 2000; Clapauch et al., 2002; Chandeying and Lamlertkittikul, 2007).

The hypothesis investigated in this study was that the steroidal saponins present in sisal juice could reverse some of the effects caused by the loss of ovarian function. Thus, the objectives of this investigation were: (a) to determine the phytochemical profile of *A. sisalana* juice and the concentration of saponins in different extracts of *A. sisalana* that have pharmacological activity; and (b) to characterise the effects of *A. sisalana* extracts on the body weight of rats in a condition that corresponds to an induced climacteric and to characterise its effects on defensive behaviours related to the manifestation of anxiety.

MATERIALS AND METHODS

Obtaining the plant materials of origin

The mucilage that was obtained by refining *A. sisalana* leaves was sourced from farmers in the city of Valente-Bahia, north-eastern Brazil (latitude: 11° 44' 24" S, longitude: 39° 27' 43" W, altitude: 358 m), through a partnership with the Syndicate of Plant Fiber Industries of the State of Bahia. The mucilage was frozen and transported to the Laboratory of Pharmaceutical Technology in Herbal Medicines of the Faculty of Sciences and Letters of São Paulo State University (UNESP), Assis. *A. sisalana* juice was acquired by pressing and filtering the mucilage.

Animals

The study was approved by the local Ethics Committee on the Use of Animals (CEUA 013/2012). All procedures were conducted in accordance with international ethical standards concerning animal experimentation.

Wistar rats, obtained from the Central Vivarium of UNESP, Botucatu, were used. The mean weight of the rats was 200 g at the beginning of the experimental sessions. The animals were housed in polypropylene cages (five animals per box). Food (Nuvilab CR-1 kibble) and water were available *ad libitum*. The vivarium was maintained with controlled temperatures (23±2°C) and with an artificial lighting programme that corresponded to 50 lux (lights on at 7:00 am and off at 7:00 pm). The animals were handled for the exchange of boxes three times per week.

*Corresponding author. E-mail: raica@assis.unesp.br.

Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> License 4.0 International License

Analysis of the chemical composition of A. sisalana juice

All phytochemical screening was performed according to the recommendations from previous study (Collins and Braga, 1988; Simões et al., 2010; Bessa et al., 2013). The tests were carried out with A. sisalana juice. The different phytochemical compounds present in this juice were characterised by performing chemical reactions that resulted in the development of foam, colouring or precipitation. The following compounds analysed and reactions used were: saponins, agitation-froth test, coumarins, reaction with aqueous potassium hydroxide solution; alkaloids, reaction with Dragendorff's reagent; flavonoids, Shinoda, Taubock, aluminium chloride and Pew tests; anthraquinones, Bornträger's reaction (direct and with prior acid hydrolysis); tannins, identification tests of hydrolysed and condensed tannins with solutions of ferric chloride. acetic acid and lead acetate, and by Stiasny's reaction. A positive or negative result was determined for each phytochemical compound based on whether the expected characteristic reaction was or was not observed.

Preparation of A. sisalana extracts

In this study, concentrated crude juice (CCJ) was obtained by heating *A. sisalana* juice to 100°C and reducing the volume six-fold. In addition, three extracts from the mucilage of *A. sisalana* were developed: (1) dried extract (DE) that resulted from centrifugation (Fanem FR 22) of the natural juice at 4000 rpm for 20 min; (2) dry mucilage extract (DME) was obtained after mucilage was dried in the sun for a week then crushed, rehydrated to a smaller volume, pressed, filtered and centrifuged at 4000 rpm for 20 min; (3) intermediate product (IP) was obtained from *A. sisalana* juice that was initially subjected to sulphuric acid to obtain a pH of 0.4 to 0.8, heated to a high temperature (120°C) for 150 min and centrifuge (Fanem FR 22) at 4000 rpm for 20 min. All extracts were dried in a greenhouse (Fanem 002CB model) at 50°C until a constant weight was achieved.

Because the glycosidic portion of saponins makes these large molecules water soluble (Simões et al., 2010), all extracts were dissolved in distilled water. However, considering that chemical hydrolysis of the IP extract is able to remove the sugar chain (the hydrophilic portion of saponins), the lipophilic portion or the steroidal sapogenins (Bodeiko and Kintya, 1975), it was necessary to dissolve in 2% Tween 80 mixed with distilled water.

Application of A. sisalana extracts

CCJ (500 or 1000 mg/kg), DE (50, 100 or 200 mg/kg), DME (25, 50 or 100 mg/kg) and IP (25, 50 or 100 mg/kg) were administered through the orogastric pathway using a syringe attached to a gavage probe. Doses were administered in the morning in a volume of 2 mL per animal. The same procedure was followed for animals of the control group (sham and ovariectomized rats); they received only the vehicle (distilled water) in the same way as the treated animals and with the same volume. Another control group was designed in which the rats were treated with distilled water plus 2% Tween 80 because the IP was prepared with this substance. The animals were weighed three times per week, immediately before the morning dose administration, to ensure appropriate dosage. The doses were defined based on previous results obtained by a research group in other studies (unpublished).

Study description

After arriving from the central vivarium, the rats were grouped (five animals per box) and left for a week to acclimatise. The treatment,

as described previously, was started after the one-week acclimatisation period, and it lasted for a period of 15 days. On day 15, the rats were subjected to an ovariectomy with the goal of interrupting their hormonal cycle. The sham animals underwent a surgical procedure, which was identical to those of the treatment animals except for removal of the ovaries. Twenty-one days after the procedures, behavioural assessments were performed (on day 36 after the first application of the extract or control solution).

Two experiments were developed. The first experiment was designed to evaluate the effect of A. sisalana CCJ (CCJ 500 or 1000 mg/kg) in comparison to a control group of females that underwent sham surgery and ovariectomized rats treated only with distilled water (2 mL/animal) (sham, n = 15; ovariectomized, n = 15; CCJ 500 mg/kg, n = 15; and CCJ 1000 mg/kg, n = 14).

The second experiment was designed to evaluate the effects of different preparations and their controls. The experimental groups were: Sham females that received daily doses of distilled water (2 mL/animal), n = 10; ovariectomized rats that received daily doses of distilled water (2 mL/animal), n = 18; ovariectomized + Tween 80, rats that received daily doses of distilled water plus 2% Tween 80 (2 mL/animal), as a parameter to the IP that was diluted with the aid of this emulsifier and surfactant, n = 9; DE 50 mg/kg, n = 9; DE 100 mg/kg, n = 10; DE 200 mg/kg, n = 8; DME 25 mg/kg, n = 9; DME 50 mg/kg, n = 10; DME 100 mg/kg, n = 10; IP 25 mg/kg, n = 10; IP 50 mg/kg, n = 7; and IP 100 mg/kg, n = 9. All groups received doses or volumes in a single application (during the morning) for 36 days, as previously described.

Behavioural assessment

The tests were performed under low illumination, similar to the vivarium lighting (50 lux). Other environmental conditions were exactly the same as the vivarium. The tests were recorded through a video system and analysed using the EthoLog 2.25 program (Ottoni, 2000).

The animals were evaluated in an EPM test, which allows for an assessment anxiety. It was elevated 50 cm from the floor and it consisted of two open arms (50 \times 10 cm) and two closed arms (50 \times 10 \times 40 cm) that are arranged in such a way that the closed arms were perpendicular to the open arms (Pellow et al., 1985). To avoid animals falling off, the edges of the open arms were protected by wood that measured 3 mm high and 2 mm thick. Animals were assessed for a period of 5 min. The results were expressed as the percentage of entries and time in open arms in relation to total entries and time in open and closed arms, respectively. The number of entries in the closed arms was measured as an index of motor activity. Entrance into an arm or across the centre was considered to have occurred with the complete passage of all four legs.

Immediately after being tested in the EPM, the animals were evaluated in a 60 x 60 cm wooden arena (open field) that was made up of 20 x 20 cm demarcated squares (20 \times 20 cm) for a period of 5 min. The animals were initially placed in the central quadrant of the box. In order to check if the motor activity of the tested animals influenced the behavioural test, the number of quadrants traversed by the animals was counted. An animal was counted as having crossed a square if it crossed the intersection of the lines with all four legs.

After the behavioural evaluations, each animal was carefully anesthetised and subsequently sacrificed. Some organic structures were removed and weighed, such as, spleen, adrenals, thymus, uterine tubes and uterus.

Body weight

Animals were weighed through the duration of the experiment in

order to check possible differences in weight variation between groups. For this, the differential weight of animals were used, which was the difference between the final weight (observed on the day of behavioural evaluation) and starting weight (noted at the beginning of treatment) of each animal.

Detection of total saponin concentrations

The concentration of saponins in *A. sisalana* extract that presented the best result was evaluated using a spectrophotometric analysis, according to the recommendations from previous study (Clark et al., 1993; Vigo et al., 2003). The analysis was performed in a UV-Vis spectrophotometer. The *A. sisalana* dried extract was dissolved in distilled water at three concentrations: 0.25, 0.35 and 0.50 mg/mL. An aliquot of 1 mL of each extract solution was added to 1 mL of 0.2% cobalt chloride chromogenic reagent and 1 mL concentrated sulphuric acid. To verify the absorbance, the reading of the solution occurred at 284 nm 20 min after the start of the reaction. In this analysis, the saponin (Merck, 0.2 mg/mL) was used as a positive control.

The concentration of total saponins was expressed in milligram of saponins per milliliter of extract solution and in milligram of saponins per gram of the dry extract. For the calculation of these concentrations, a linear regression curve was used to establish a straight line equation, which was then applied at concentrations of 0.08 to 0.28 mg/mL.

Data analysis

The behavioural data in each experiment, as well as the body weight, the weight of organic structures (such as spleen, adrenals, thymus, uterine tubes and uterus) and the consumption of water and food were analysed by an analysis of variance (ANOVA), followed by Duncan's post hoc test. In all cases, results with a p value less than or equal to 0.05 were considered significant.

RESULTS

Phytochemical analysis

The tests for detecting and prospecting the chemical constituents of A. sisalana juice indicated the presence of tannins saponins, coumarins, flavonoids, and anthraguinones (Table 1). A high saponin content in A. sisalana juice was detected by observing foam permanence, even after the addition of hydrochloric acid. The presence of coumarin was detected by the reaction of this compound with potassium hydroxide. In the search for flavonoid, using metallic magnesium powder and concentrated hydrochloric acid, the presence chalcones, aurones, dihydrochalcones and isoflavones in A. sisalana juice were identified by the cyanidin or Shinoda reaction. The presence of flavones, flavonols and flavonones occurred by a reaction of A. sisalana juice with aluminium chloride, and flavanones and isoflavones with the boric acid and oxalic acid solutions. also known as reaction Taubock. Using Pew's reaction, no anthocyanins were detected in the A. sisalana juice. Ferric chloride and Stiasny's reaction were used for detection of condensed tannins. The presence of hydrolysable tannins was not detected. Usina Bornträger's direct reaction, the presence of free anthraquinones was determined by observing a red colouration in the region that made contact with the juice and by observing a brown colour after homogenisation. Bornträger's reaction with prior acid hydrolysis was used to identify anthraguinones, glycosides and dimers. The presence of these types of anthraguinone in the juice was not identified because the reaction did not result in a rosy-coloured reaction, which is indicative of a positive result. The formation of precipitates through direct survey in the samples was not observed, suggesting the absence of alkaloids.

Behavioural and body weight analysis

Experiment 1: Effect of treatment with A. sisalana CCJ (500 or 1000 mg/kg) on the manifestation of defensive behaviours, body weight and some structures of ovariectomized females

The two doses of *A. sisalana* CCJ used in this study did not cause behavioural changes in the EPM. Specifically, a statistical change was not observed in the percentage of entries in open arms/total [F (3.54) = 1.07; p = 0.368]; the percentage of time in open arms/total [F (3.54) = 1.34; p = 0.272] (Figure 1); or entries in closed arms [F (3.54) = 0.88; p = 0.458] or in the arena [F (3.54) = 1.58; p = 0.204] (Figure 2). Also, there were no alterations observed in the weight of organic structures (such as spleen, adrenals, thymus, uterine tubes and uterus), change in the amount of water or change in the amount of food ingested (p > 0.05). However, there was a significant observation of a loss of body weight [F (3.55) = 28.26; p < 0.001] (Figure 3).

Experiment 2: Effect of treatment with different preparations of A. sisalana (DE, DME or IP) on the manifestation of defensive behaviours, body weight and some structures of ovariectomized females

In the EPM, an analysis of the percentage of entries in open arms/total, ANOVA showed that in relation to different doses of DE, DME and IP, the result was not significant [F (11.11) = 1.87; p = 0.051]. However, considering that the result was borderline, Duncan's test showed the ovariectomized females treated with DE at 50 and 200 mg/kg, DME at 100 mg/kg and IP at 50 mg/kg IP explored the open spaces of the EPM less than the control group females (ovariectomized + Tween 80; p < 0.01). For the percentage of time in open arms/total, ANOVA showed that there were no differences in a comparison of the effects of treatments at different doses of DE, DME and IP [F (11.11) = 1.20; p = 0.292]. Figure 1 illustrates these results.

Table 1. Phytochemical prospecting tests of the sisal juice.

Chemical constituent	Positive reaction	Result observed
Saponins	Stable foam layer for more than 30 m	Positive
Coumarins	Blue fluorescence under ultraviolet light	Positive
Flavonoids	Reaction with aluminium chloride; yellow fluorescence under ultraviolet light for flavones and flavonols; blue-green for flavanones	Positive for flavones, flavonols and flavonones
	Shinoda's reaction: Yellow to red for flavones; red to bloody red for flavanols and dihydroflavonols; red to purple for flavanones; and red to pink for anthocyanic derivatives Negative reaction without staining for chalcones, aurones, dihydrochalcones and isoflavones	The reaction was colourless, indicating the presence of chalcones, aurones, dihydrochalcones and isoflavones
	Pew's reaction: Reddish for anthocyanins	Negative
	Taubock's reaction: Yellow fluorescence under ultraviolet light for flavonols. Flavones and isoflavones do not present fluorescence. Anthocyanic compounds stain but do not produce fluorescence	The reaction did not present fluorescence, indicating the presence of flavones and isoflavones
Tannins	Stiasny's reaction: Red precipitate for condensed tannins and blue precipitate for hydrolysable tannins	Red precipitate for condensed tannins
	Reaction with ferric chloride: Blue for hydrolysable tannins and green for condensed tannins	Green colouration, indicating the presence of condensed tannins
	Reaction with acetic acid and acetate of lead: Whitish precipitate for hydrolysable tannins	Negative
Anthraquinones	Bornträger's direct reaction: Red tinge in the area in contact with the juice for free anthraquinones (brown when homogenised)	Positive
	Bornträger's reaction with prior acid hydrolysis: Rosy colouring to identify anthraquinones, glycosides and dimers	Negative
Alkaloids	Reddish-orange precipitate	Negative

In relation to entry in the closed arms (Figure 2), there were significant effects between the treatments and their respective doses of *A. sisalana* [F (11.11) = 3.24; p < 0.001], as can be seen in Figure 2. Based on results from Duncan's test, rats that received doses of 100 mg/kg of DME (p < 0.01) showed a significant increase in activity in relation to the control group (ovariectomy + Tween 80).

For the number of squares covered in the arena (Figure 2), there was an effect between the treatments and their respective doses of A. sisalana [F (11.11) = 3.94; p < 0.001]. Results from Duncan's test demonstrated that ovariectomized rats treated with distilled water + Tween 80 showed a significant increase in motor activity compared to ovariectomized rats and those treated only with distilled water (p < 0.05). Rats treated with DE 50, 100 and 200 mg/kg showed a decrease in motor activity only when compared to the control group (ovariectomized + Tween 80; p < 0.05). Results were also different for rats treated with IP (25 to 100 mg/kg), which showed an increase in motor activity compared to the specific

control group, especially those treated with DE 50 mg/kg.

When the mean differential variation of weight was analysed by ANOVA (Figure 3), the results showed that there was an effect between the treatments and their respective doses of *A. sisalana* [F (11.11) = 8.45; p < 0.001]. Duncan's test indicated that ovariectomy caused an increase in body weight. Rats treated with DE at 100 and 200 mg/kg, loss weight when compared with control rats (ovariectomized with or without Tween) (p < 0.001). Rats treated with DE at 50 mg/kg were only different from ovariectomized + Tween 80 rats (p < 0.05), compared to the control group (ovariectomized + Tween 80).

Because the rats treated with DE at 100 mg/kg did now show any changes in behaviours in the EPM or open field tests compared to the rats that were only ovariectomized, other variables were analysed. It was found that this extract, DE, caused an increase in the weight of the reproductive structures and of the adrenals (p < 0.01), but it did not cause changes in the weight of the other organs, such as the spleen and liver, or a change in the

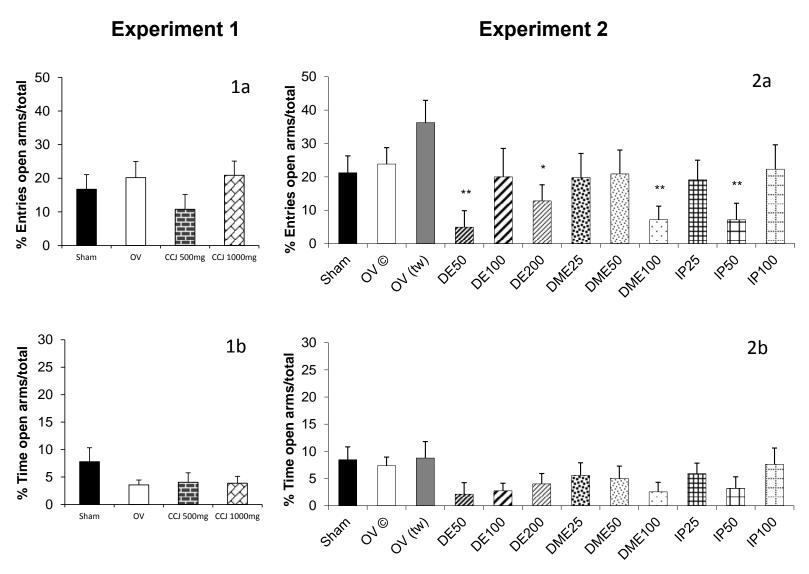


Figure 1. Manifestation of defensive behaviours related to anxiety in the elevated plus maze. Mean + standard error of the mean (SEM) of the percentage of entries and time in open arms in relation to total entries and time in open and closed arms, respectively. On the left are data from Experiment 1 (1a: % entries/total; 1b: % time/total): treatment with *Agave sisalana* concentrated crude juice (CCJ, 500 or 1000 mg/kg) and control group. On the right are data from Experiment 2 (Panel 2a: % entries/total; Panel 2b: % time/total): Different doses of the dried extract (DE), dry mucilage extract (DME) and intermediate product (IP) of *A. sisalana* and control groups. A p-value of less than 0.05 (*) and less than 0.01 (**) represents the significant differences in relation to the control group of ovariectomized + Tween 80 (analysis of variance followed by Duncan's post hoc test).

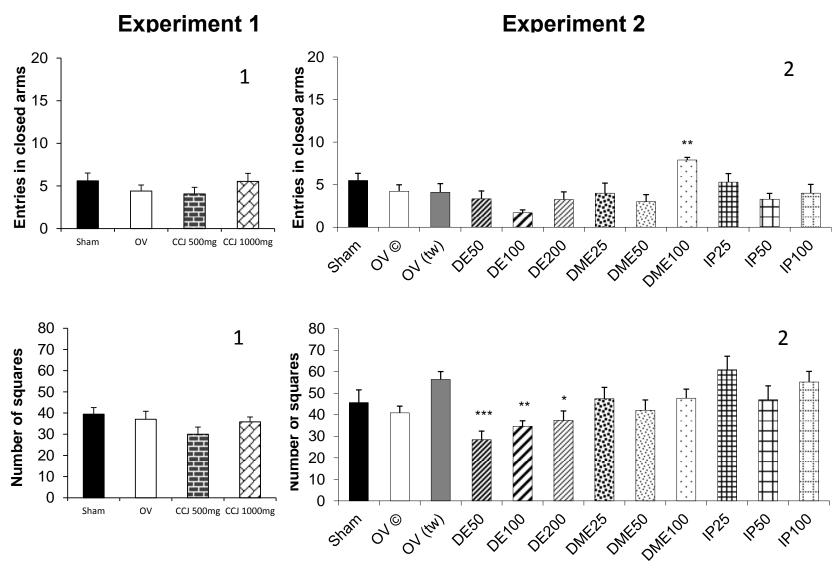
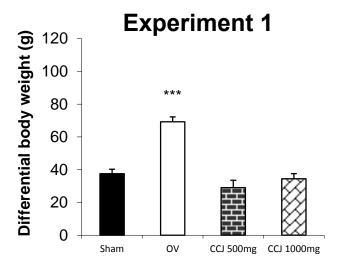


Figure 2. Motor activity of females in the elevated plus maze and the arena. Mean + standard error of the mean (SEM) of the percentage of entries and entries in open arms in relation to total entries and time in open and closed arms, respectively. On the left are data from Experiment 1 (1a: % entries/total; 1b: % time/total): *A. sisalana* concentrated crude juice - CCJ, 500 or 1000 mg/kg groups and control group. On the right are data from Experiment 2 (2a: % entries/total; 2b: % time/total): different doses of dried extract (DE), dry mucilage extract (DME) and intermediate product (IP) of *A. sisalana* and control groups. A p-value of less than 0.05 (*) and less than 0.01 (**) represents the significant differences in relation to the control group of ovariectomized + Tween 80 (analysis of variance followed by Duncan's post hoc test); p > 0.05, result not significant (ANOVA followed by Duncan's post hoc test).



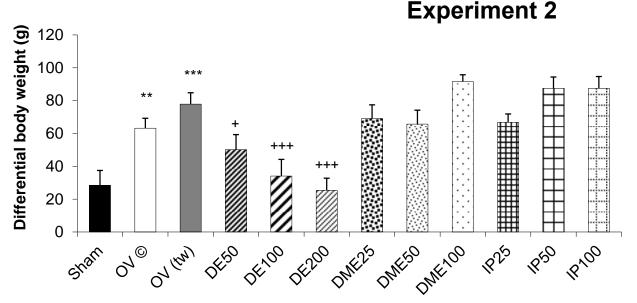


Figure 3. Differential body weight of females (change from initial weight measurement to final weight measurement). Mean + standard error of the mean (SEM) of the differential body weight of ovariectomized females. On the left are data from Experiment 1: *A. sisalana* concentrated crude juice - CCJ, 500 or 1000 mg/kg groups and control group. On the right are data from Experiment 2: Different doses of dried extract (DE), dry mucilage extract (DME) and intermediate product (IP) of *A. sisalana* and control groups (analysis of variance followed by Duncan's post hoc test). A p-value of less than 0.05 (*) and less than 0.001 (**) represents the significant differences in relation to the control group of ovariectomized + Tween 80.

amount of water and food ingested (p > 0.05).

Detection of total saponin concentrations

Because three different concentrations of DE were associated with a decrease in body weight in ovariectomized rats without affecting behaviour in the EPM or in the arena, this extract was analysed to determine the concentration of total saponins. To calculate the concentration of total saponins present in the three concentrations (Table 2), a straight equation

(y=1.9841x + 0.0995) was established using the curve of a linear regression with the saponin (Merck). In the straight equation, x corresponds to the analysed concentration of saponins (mg/ml) and y corresponds to the absorbance measured. A line obtained with R2=0.9963, confirms the linearity of the reading of the concentrations that were used.

In addition, starting with the dilution factor of the DE solution analysed, the concentration of total saponins in "milligram" per "gram" of the dry extract was also expressed. The average of the values obtained in this analysis showed a high concentration of total saponins

Table 2. Values of total saponins presented in dried extract.

Concentration of extract (mg/mL)	Absorbance	Concentration of saponins (mg/mL)*	Concentration saponins (mg/g)**
0.25	0.537	0.22	882.68
0.35	0.737	0.32	918.01
0.50	1.017	0.46	924.52

^{*}Results were expressed in milligram of total saponins per milliter of dried extract solutions analysed. **Results were expressed in milligram of total saponins per gram of the dried extract.

(908.40 mg/g) in this extract; this was a proportion of 90.84% of the weight of the dry extract.

For more assurance about the accuracy of the obtained data, 0.2 mg/mL of the saponin (Merck) was used as positive control for the analysis of the values of absorbance of the DE solutions; an absorbance of 0.471 was reported. For comparison purposes, using the straight equation (y=1.9841x + 0.0995), a value of 0.496 was obtained for the absorbance of 0.2 mg/mL of saponin, which confirmed the precision of the method used.

DISCUSSION

The phytochemical study of A. sisalana juice revealed the presence of a diverse group of secondary metabolites. including coumarins, flavonoids, condensable tannins, alkaloids, free anthraquinones and mainly saponins. These secondary metabolites are described as having many biological properties and therapeutic actions. Emphasis should be given to the antioxidant, antiinflammatory and antitumour activities of flavonoids (Machado et al., 2008; Simões et al., 2010) and especially to the biological properties of saponins, whose presence was largely detected in this preliminary phytochemical study. Saponins have numerous activities, such as antifungal, anti-allergic, anti-inflammatory, antibiotic and antitumour activities (Francis et al., 2002; Sparg et al., 2004). Furthermore, saponins exhibit an anxiolytic effect in the EPM (Wei et al., 2007), and they are able to interfere in the metabolism of cholesterol and fat absorption by inhibiting pancreatic lipase activity (Dickel et al., 2007).

Based on the biological properties associated with secondary metabolites, especially the saponins, it has been suggested that *A. sisalana* juice could contribute to the pharmacological effects related to the objectives of this investigation (that is, the behavioural and physiological assessments).

In this regard, ovariectomy, as expected, caused an increase in the body weight of females in both experiments (Chen and Heiman, 2001), and treatment with two different concentrations of CCJ caused weight loss in ovariectomized rats. Similarly, rats treated with DE

(50, 100 or 200 mg/kg) also demonstrated a reduction in body weight.

Contrary to observations reported previously (Lagunas et al., 2010), the ovariectomy did not cause any behavioural modification in the EPM or open field tests. However, considering that many pharmacological treatments of obesity cause several adverse effects to the body, including an increase in anxiety and in general activity (Ioannides-Demos et al., 2011; Carter et al., 2012; Haslam, 2016; Nuffer et al., 2016), it has become of extreme importance to verify if the preparations of *A. sisalana* that cause weight loss modify the anxiety profile or cause motor disorders.

A. sisalana CCJ did not modify the behavioural responses of ovariectomized rats in the EPM and open field tests, which indicate that it is a potent candidate for the treatment of weight gain that is associated with menopause. Therefore, it is important to highlight the results obtained with DE at doses of 100 and 200 mg/kg because those doses promoted a reduction in body weight that was acquired after an ovariectomy, and it did not affect the manifestation of behaviour related to anxiety and motor activity when compared with ovariectomized rats treated only with distilled water (without Tween). In addition, the treatment with DE at 100 mg/kg caused an increase in the weight of the structures of the reproductive system.

The presence of saponins in *A. sisalana*, CCJ, and especially in DE extract, may explain the observed reduction in body weight. In fact, the spectrophotometric analysis of DE showed a high proportion of total of saponins (90.84%) in relation to the weight of the dry extract. Thus, the reduction in body weight observed in ovariectomized rats and the increase in the weight of the reproductive system structures seem to be related to the presence of saponins. According to Francis et al. (2002), the effects of saponins appear to be related to interactions with steroid receptors, once the basic structures of saponins are similar to steroidal hormones.

As already mentioned, studies with plants used for weight loss reported that the saponins in these plants might be able to interfere in the metabolism of cholesterol and fat absorption by inhibiting pancreatic lipase activity (Dickel et al., 2007). Natural and synthetic saponins inhibit the absorption of cholesterol in the intestine,

reduce the concentration of plasma cholesterol in laboratory animals and are used for their pharmacological potential in the treatment of hypercholesterolaemia (Harwood Jr. et al., 1993). In recent studies, Liu et al. (2015) reported that the dioscin, a natural steroid saponin that exists widely in various agave plants (Sidana et al., 2016), could cause gradual weight loss without inhibiting appetite or increasing the physical activity of obese mice. In addition, Leal-Díaz et al. (2016) demonstrated that steroidal saponins extracted from Agave salmiana were able to reduce obesity-related metabolic abnormalities by promoting an abundance of Akkermansia muciniphila in the intestinal lumen. The presence of these bacteria is inversely associated with insulin resistance, altered adipose tissue metabolism, the onset of inflammation and obesity development during diet-induced obesity in mice (Schneeberger et al., 2015).

Besides that, in toxicity studies, any change in the weight of reproductive or non-reproductive organs may be a good indication of toxicity that is promoted by one or more phytochemical compounds from medicinal plants. On the basis of this information, a toxicological study was carried out by a research group in adult female Wistar rats using a hydrolysed extract obtained from A. sisalana (unpublished). Results show that oral treatment (gavage) for 30 consecutive days promoted a slight increase in the absolute weight of ovarian tissues; however, the relative weight was unaltered in the group administered sisal juice. The uterus, heart, liver and kidney weights were not affected by exposure to A. sisalana extract. Furthermore, the plant did not cause mortality in experimental groups, and it did not promote signs of toxicity such as changes in behaviour, ataxia, salivation, vomiting, diarrhoea, polyphagia, fever, weakness, tremors or convulsions (Viel et al., 2017). In addition, a lack of toxicity was observed when the hexane fraction obtained from the hydrolysed extract of A. sisalana was administered in an acute dose to rats and mice; the chemical high-performance liquid chromatography analyses of the hexane fraction of A. sisalana confirmed the presence of two steroidal sapogenins, hecogenin and tigogenin (Dunder et al., 2013).

The results obtained in this study are of high clinical relevance. This is because one of the major problems associated with the loss of ovarian function, whether by ovary removal surgery or functional loss resulting from natural ageing in women (menopause), is body weight gain. In addition to involving health problems such as an increase in rates of cholesterol (low-density lipoprotein) that can lead to cardiovascular diseases, weight gain can severely affect a woman's self-esteem, increase anxiety and manifest as depression in this stage of life.

Conclusions

Together, the results obtained indicate that chronic treatment with *A. sisalana* CCJ and with DE (100 mg/kg)

caused a decrease in body weight without affecting behavioural manifestations related to anxiety or motor alteration, but with relevant stimulation of the structures of the reproductive system. As an expressive concentration of saponins was identified in DE, the effects observed were attached to this component. Such findings point to *A. sisalana* as a potential product in the treatment of weight gain in menopause, as it did not interfere with the manifestation of defensive behaviours, and it also brought on stimulation of internal structures of the reproductive system as a potent phytoestrogen. To find a compound that causes weight loss without an increase in anxiety would be a major breakthrough, which strengthens the relevance of this investigation.

CONFLICT OF INTERESTS

The authors have not declared any conflict of interests.

ACKNOWLEDGMENTS

This work was supported by grant #2010/18478-6 from São Paulo Research Foundation (FAPESP) and by the Syndicate of Plant Fiber Industries of the State of Bahia.

REFERENCES

Attele AS, Zhou YP, Xie JT, Wu JA, Zhang L, Dey L, Pugh W, Rue PA, Polonsky S, Yuan CS (2002). Antidiabetic effects of Panax ginseng berry extract and the identification of an effective component. Diabetes 51(6):1851-1858.

Azevedo AIB, Miranda JE, Soares JJ, Souza Júnior JDA, Moreira MD (2003). Efeitos do extrato de sisal (Agave sisalana) sobre o curuquerê (*Alabama argillacea*) (Lepidoptera: noctuidae). In: Congresso Brasileiro de Algodão, 4, Goiânia. Proceedings,Goiânia. Available

http://www.cnpa.embrapa.br/produtos/algodao/publicacoes/trabalhoscba4/069.pdf

Bessa NGF, Borges JCM, Beserra FP, Carvalho RHA, Pereira MAB, Fagundes R, Campos SL, Ribeiro LU, Quirino MS, Chagas Junior AF, Alves A (2013). Prospecção fitoquímica preliminar de plantas nativas do cerrado de uso popular medicinal pela comunidade rural do assentamento vale verde - Tocantins. Rev. Bras. Plantas Med. 15(4):692-707.

Bodeiko A, Kintya K (1975). The structure of Agave saponins C' and D from the leaves of Agave americana. Chem. Nat. Compd. 11(6):775-777

Bonduki CE, Haidar MA, Lima GR, Baracat EC (2006). Terapia de reposição hormonal em mulheres na pós-menopausa. Psiq. Prat. Med. 24(1):172-181.

Bown D (1995). The Royal Horticultural Society encyclopedia of herbs and their uses. Dorling Kindersley Limited.

Carter R, Mouralidarane A, Ray S, Soeda J, Oben J (2012). Recent advancements in drug treatment of obesity. Clin. Med. 12(5):456-460.

Chandeying V, Lamlertkittikul S (2007). Challenges in the conduct of Thai herbal scientific study: Efficacy and safety of phytoestrogen, puerariamirifica (KwaoKeur Kao), phase I, in the alleviation of climacteric symptoms in perimenopausal women. J. Med. Assoc. Thai. 90:1274-1280.

Chen Y, Heiman ML (2001). Increased weight gain after ovariectomy is not a consequence of leptin resistance. Am. J. Physiol. Endocrinol. Metab. 280(2):E315-E322.

- Clapauch R, Meirelles RMR, Julião MASG, Loureiro CKC, Giarodoli PB, Pinheiro SA, Harringan AR, Spritzer PM, Pardini DP, Weiss RV, Athayde A, Russo LA, Póvoa LC (2002). Fitoestrogênios: Posicionamento do Departamento de Endocrinologia Feminina da Sociedade Brasileira de Endocrinologia e Metabologia (SBEM). Arq. Bras. Endocrinol. Metab. 46(6):679-695.
- Clark BJ, Frost T, Russel MA (1993). Techniques in visible and ultraviolet spectrometry. London: Chapman and Hall. 146p.
- Collins CH, Braga GL (1988). Introdução a métodos cromatográficos. 3rd ed. Campinas: Unicamp. P 452.
- Dickel ML, Rates SM, Ritter MR (2007). Plants popularly used for loosing weight purposes in Porto Alegre, South Brazil. J. Ethnopharmacol. 109(1):60-71.
- Duke JA, Ayensu ES (1985). Medicinal plants of China. Reference Publications. Vol. 2.
- Dunder RJ, Luiz-Ferreira A, Almeida ACAD, de-Faria FM, Takayama C, Socca EAR, Salvador MJ, Mello GC, Santos C, Oliva-Neto P, Souza-Brito ARM (2013). Applications of the hexanic fraction of *Agave* sisalana Perrine ex Engelm (Asparagaceae): control of inflammation and pain screening. Mem. Inst. Oswaldo Cruz. 108:263-271.
- El-Hilaly J, Hmammouchi M, Lyoussi B (2003). Ethnobotanical studies and economic evaluation of medicinal plants in Taounate province (Northern Morocco). J. Ethnopharmacol. 86(2-3):149-58.
- Francis G, Kerem Z, Makkar HP, Becker K (2002). The biological action of saponins in animal systems: a review. Br. J. Nutr. 88(6):587-605.
- Han LK, Xu BJ, Kimura Y, Zheng Y, Okuda H (2000). Platycodi radix affects lipid metabolism in mice with high fat diet-induced obesity. J. Nutr. 130(11):2760-2764.
- Harwood Jr HJ, Chandler CE, Pellarin LD, Bangerter FW, Wilkins RW, Long CA, Cosgrove PG, Malinow MR, Marzetta SCA, Pettini JL, Savoy YE, Mayne JT (1993). Pharmacologic consequences of cholesterol absorption inhibition: alteration in cholesterol metabolism and reduction in plasma cholesterol concentration induced by the synthetic saponin P-tigogen in cellobioside. J. Lipid Res. 34:377-395.
- Haslam D (2016). Weight management in obesity- past and present. Int. J. Clin. Pract. 70(3):206-217.
- Hostettmann K, Marston A (1995). Chemistry and Pharmacology of Natural Products, Saponin. Cambridge University Press, Cftmbridge.
- Ioannides-Demos LL, Piccenna L, McNeil JJ (2011). Pharmacotherapies for obesity: past, current, and future therapies. J. Obes. 2011:1-18.
- Lagunas N, Calmarza-Font I, Diz-Chaves Y, Garcia-Segura LM (2010). Long-term ovariectomy enhances anxiety and depressive-like behaviors in mice submitted to chronic unpredictable stress. Horm. Behav. 58:786-791.
- Leal-Díaz AM, Noriega LG, Torre-Villalvazo I, Torres N, Alemán-Escondrillas G, López-Romero P, Sánchez-Tapia M, Aguilar-López M, Furuzawa-Carballeda J, Velázquez-Villegas LA, Avila-Nava A, Ordáz G, Gutiérrez-Uribe JA, Serna-Saldivar SO, Tovar AR (2016). Agave salmiana and its extracted saponins attenuated obesity and hepatic steatosis and increased Akkermansia muciniphila in C57BL6 mice. Sci. Rep. 6:34242.
- Lima GR, Baracat EC (1995). Síndrome do climatério. In: Lima GR, Baracat EC, eds. Ginecologia endócrina. 1st ed. São Paulo: Editora Atheneu. pp. 253-257.
- Liu M, Xu L, Yan L, Youwei X, Han X, Zhao Y, Sun H, Yao J, Lin Y, Liu K, Peng J (2015). Potent effects of dioscin against obesity in mice. Sci. Rep. 5:7973.
- Machado H, Oliveira TT, Nagem TJ, Peters VM, Fonseca CS (2008). Flavonoids and potential therapeutic. Boletim Centro Biol. Reprod. Univ. Fed. Juiz Fora 27:33-39.
- Martin AR, Martins MA, Mattoso LHC, Silva ORRF (2009). Caracterização química e estrutural de fibra de sisal da variedade Agave sisalana. Polímeros 19(1):40-46.
- Martin AR, Denes FS, Rowell RM, Mattoso LHC (2003). Mechanical behavior of cold plasma–treated sisal and high-density polyethylene composites. Polym. Compos. 24(3):464-474.
- Milewicz A, Jedrzejuk D (2006). Climacteric obesity: From genesis to clinic. Gynecol. Endocrinol. 22(1):18-24.
- Nuffer W, Trujillo JM, Megyeri J (2016). A comparison of new pharmacological agents for the treatment of obesity. Ann. Pharmacother. 50(5):376-388.

- Oashi MCG (1999). Estudo da cadeia produtiva como subsídio para pesquisa e desenvolvimento do agronegócio da *Agave sisalana* na Paraíba. Doutorado em Engenharia de Produção. Thesis. Universidade Federal de Santa Catarina, Florianópolis.
- Ottoni EB (2000). EthoLog 2.2: A tool for the transcription and timing of behavior observation sessions. Behav. Res. Methods Instrum. Comput. 32(3):446-449.
- Pellow S, Chopin P, File SE, Briley M (1985). Validation of open-closed arm entries in an elevated Plus-Maze as a measure of anxiety in the rat. J. Neurosci. Methods 14(3):149-167.
- Pizarro AB, Oliveira Filho AM, Parente JP, Melo MTV, Santos CE, Lima PR (1999). O aproveitamento do resíduo da indústria do sisal no controle de larvas de mosquitos. Rev. Soc. Bras. Med. Trop. 32(1):23-29.
- Popov AA, Izmozherov NV, Tagil'tseva NV, Andreev AN, Striukova O, Fominykh MI, Akimova AV (2007). Osteoarthrosis of the knee joints in climacteric womem with excessive body weight or obesity. Klin. Med. (Mosk). 85(9):64-67.
- Rachev E, Stamenov G, Davidkova, N (2000). Influence of the phytoestrogen drug, SoyaVital, on climacteric symptoms. Akush. Ginekol. 40(4):51-52.
- Ribeiro BD, Alviano DS, Barreto DW, Coelho MAS (2013). Functional properties of saponins from sisal (*Agave sisalana*) and juá (*Ziziphus joazeiro*): critical micellar concentration, antioxidant and antimicrobial activities. Colloids Surf. A. Physicochem. Eng. Asp. 436:736-743.
- Schneeberger M, Everard A, Gómez-Valadés AG, Matamoros S, Ramírez S, Delzenne NM, Gomis R, Claret M, Cani PD (2015). Akkermansia muciniphila inversely correlates with the onset of inflammation, altered adipose tissue metabolism and metabolic disorders during obesity in mice. Sci. Rep. 5:16643.
- Sclowitz IKT, Santos IS, Silveira MF (2005). Prevalência e fatores associados a fogachos em mulheres climatéricas e pós-climatéricas. Cad. Saúde Pública. 21(2):469-481.
- Sharaf A, Zahran M (1967). Pharmacological investigation on Agave sisalana perr., with special study of its ecbolic effect. Plant Food Hum. Nutr. 14(4):345-351.
- Sidana J, Singh B, Sharma OP (2016). Saponins of Agave: Chemistry and bioactivity. Phytochemistry 30:22-46.
- Silva ORRF, Beltrão EM (1999). O Agronegócio do sisal no Brasil. Brasília: Embrapa-SPI/ Embrapa-CNPA.
- Silva RM, Araújo CB, Silva AR (2003). Alterações biopsicossociais da mulher no climatério. Rev. Bras. Promoç. Saúde. 16(1/2):28-33.
- Simões CMO, Schenkel EP, Gosmann G, Mello JCP, Mentz LA, Petrovick PR (2010). Farmacognosia: da planta ao medicamento. 6 ed. Porto Alegre/Florianópolis: UFRGS/ EDUFSC.
- Simons V, Morrissey JP, Latijnhouwers M, Csukai M, Cleaver A, Yarrow C, Osbourn A (2006). Dual effects of plant steroidal alkaloids on Saccharomyces cerevisiae. Antimicrob. Agents Chemother. 50(8):2732-2740.
- Sparg SG, Light ME, Van Staden J (2004). Biological activities and distribution of plant saponins. J. Ethnopharmacol. 94(2-3):219-243.
- Toth MJ, Tchernof A, Sites CK, Poehlman ET (2000). Menopause-related changes in body fat distribution. Ann. N. Y. Acad. Sci.904:502-506.
- Vigo CLS, Narita E, Marques LC (2003). Validação da metodologia de quantificação espectrofotométrica das saponinas de *Pfaffia glomerata* (Spreng.) Pedersen Amaranthaceae. Rev. Bras. Farmacogn. 13(2):46-49.
- Viel AM, Pereira AR, Neres WE, Dos Santos L, Oliva Neto P, Souza EB, Silva RMG, Camargo ICC (2017). Effect of *Agave sisalana* Perrine extract on the ovarian and uterine tissues and fetal parameters: comparative interventional study. Int. J. Multispeciality Health 3(5):129-138.
- Wei XY, Yang JY, Wang JH, Wu CF (2007). Anxiolytic effect of saponins from Panax quinquefolium in mice. J. Ethnopharmacol. 111(3):613-618.
- Yun JW (2010). Possible anti-obesity therapeutics from nature–a review. Phytochemistry 71(14-15):1625-1641.
- Zhao HL, Sim JS, Shim SH, Ha YW, Kang SS, Kim YS (2005). Antiobese and hypolipidemic effects of platycodin saponins in dietinduced obese rats: evidences for lipase inhibition and calorie intake restriction. Int. J. Obes. 29(8):983-990.

Journal of Medicinal Plant Research

Related Journals Published by Academic Journals

- African Journal of Pharmacy and Pharmacology
- Journal of Dentistry and Oral Hygiene
- International Journal of Nursing and Midwifery
- Journal of Parasitology and Vector Biology
- Journal of Pharmacognosy and Phytotherapy
- Journal of Toxicology and Environmental Health Sciences

academicJournals